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Bovine median eminence contains a factor different from gonadotropin-releasing hormone (GnRH) that increases basal luteinizing hormone (LH) secretion and potentiates GnRH-stimulated LH release. We compared the effects of hypothalamic neuropeptides on basal and GnRH-stimulated LH secretion using rat pituitary cells under static incubation conditions to determine if any of them mimics the LH-releasing activity not attributable to GnRH present in bovine median eminence extracts. Both, galanin and neurotensin $(10^{.9}-10^{.5} M)$ stimulated basal LH secretion in a dose-response manner. Galanin increased 3-4 fold and neurotensin doubled the basal LH secretion. The GnRH antagonist Nal-Glu 10⁶M abolished the effect of 10^{.7} M GnRH and 10^{.5} M neurotensin, but did not block the LH-releasing activity of galanin. Leucine-enkephalin, β -endorphin, substance P and neuropeptide Y (NPY) did not alter basal LH secretion. Neuropeptides produced three types of response on GnRH-stimulated LH release. First, leucineenkephalin and β -endorphin (10⁻⁹-10⁻⁵ M) showed a dose-dependent inhibition of GnRH-stimulated LH release. At 10⁻⁵ M the inhibition was complete with leucineenkephalin and only 30% with β -endorphin. Both were blocked by naloxone. Second, substance P showed an inverted U type response on GnRH-stimulated LH secretion. At 10.9 M this peptide potentiated the action of GnRH. This effect decreased when the dose of substance P was increased to 10^7 M and turned inhibitory at 10^{-5} M when 10^{-7} M GnRH was used. Third, galanin and NPY potentiated the effect of GnRH on LH secretion. Neurotensin had no effect on GnRHstimulated LH release. In conclusion, rat gonadotrophs present diverse responses to neuropeptides at physiological concentrations, and -apart from GnRH- galanin is most likely the other factor present in bovine median eminence extracts that stimulates LH secretion. The data lend further support to a role of galanin in the control of LH secretion.

Key words: luteinizing hormone, neuropeptides, opioid peptides, pituitary.

INTRODUCTION

It is well established that pituitary gonadotropin secretion is primarily under the control of gonadotropin-releasing hormone (GnRH), but there is increasing evidence that this process is modulated also by multiple other factors of gonadal and hypothalamic origin. In previous studies (22), we found that bovine median eminence (ME) extracts increased both basal and GnRH-stimulated luteinizing hormone (LH) secretion from dispersed pituitary cell cultures. This LH-releasing activity was not entirely attributed to

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GnRH because it differed in its immunoreactivity, it could stimulate LH release when GnRH receptors were blocked and the maximal response obtained was larger than that evoked by GnRH alone. To approach the identification of this factor, it was deemed appropriate to determine first whether or not neuropeptides known to be present in the hypothalamus, such as galanin (27), neurotensin (1, 28), neuropeptide Y (NPY) (30) and opioid peptides (19), could account for this activity. Although, opiates (5,7,19), corticotrophin-releasing hormone (CRH) (6), α melanocyte-stimulating hormone (α -MSH) (29), galanin (24), NPY (18), pituitary adenylate cyclase-activating polypeptide (PACAP) (16) and substance P (3,43) have been shown to alter LH release, a detailed comparison of their effects on LH secretion with that of ME extracts is not warranted from the data reported, because they were obtained under widely different experimental conditions.

Here we present a systematic comparison of the effects of various neuropeptides upon basal and GnRH-evoked LH secretion

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utilizing the same assay system used for the ME extracts. Dispersed rat pituitary cells in culture under static incubation conditions were used to evaluate the LH-releasing activity of the neuropeptides, their interactions with GnRH and whether the effects elicited by concentrations used compared to those attained at portal blood and/or pituitary itself.

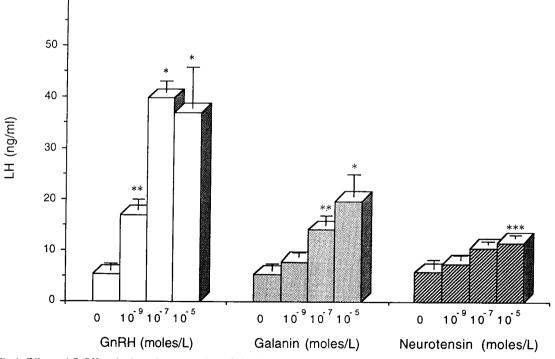
MATERIALS AND METHODS

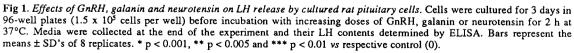
Reagents

GnRH was obtained from Bachem Bioscience Inc. The GnRH antagonist [Ac-D2Nal¹, D4ClPhe², D3Pal³, Arg⁵, D-4-p-methoxybenzoyl-2-amino butyric acid⁶, DAla¹⁰]-GnRH (Nal-Glu) was a gift from Dr Jean Rivier, Salk Institute, La Jolla, California. All other peptides, tissue culture media and reagents were obtained from Sigma Chem. Co.

Culture of rat anterior pituitary cells

Anterior pituitary glands obtained from 60days old female Sprague-Dawley rats were





dispersed as described previously (22), to obtain primary cell cultures. Dispersed cells were plated at a density of 1.5×10^5 cells per well in 96-well plates (FALCON 3872 PRI-MARIA) and cultured at 37°C in an atmosphere of 95% air-5% CO₂ for three days.

LH secretion experiments

The cells were washed twice with serum-free medium 199 (without phenol red) buffered with 10 mM HEPES, pH 7.4; containing 0.3% bovine seroalbumin (M199-BSA). The cells were then incubated for 2h at 37° C in M199-BSA alone (basal secretion) or M199-BSA containing test substances. After incubation, the media were collected for measurement of LH by ELISA. All assays were performed in eight replicates at a final volume of 0.3 ml. The effects of test substances were examined in at least two individual experiments.

LH-ELISA

A double sandwich ELISA system described

by Leiva and de la Lastra (21), was carried out in 96-well microtiter plates (Dynatech Lab. Inc.). Rat LH-RP-3 was used as standard and anti-bLH β -subunit (monoclonal antibodies, clone 518 B7 obtained from Dr Jan Roser, Department of Animal Sciences, University of California, Davis, CA.), anti-rLH-S-10 (rabbit IgG, NIDDK) and anti-rabbit IgG peroxidase conjugate (Sigma Chem Co), were used as first, second and third antibody, respectively. All samples were assayed in duplicate. Intra-assay coefficients of variation for culture medium

pools containing low, medium and high LH concentrations were 4.5, 2.3 and 6.9%, respectively. The inter-assay coefficients of variation for the same pools were 9.6, 7.3 and 8.5%, respectively.

Statistics

Values are presented as means \pm standard deviations. Significant differences were evaluated by analysis of variance (46).

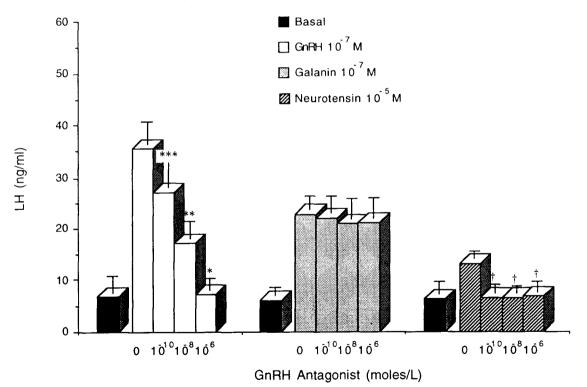


Fig 2. Effects of the GnRH antagonist, Nal-Glu, on LH-releasing activity of GnRH, galanin and neurotensin. After 3 days in culture, rat pituitary cells $(1.5 \times 10^5 \text{ cells/well})$ were preincubated in the presence of increasing doses of Nal-Glu for 30 min and then were coincubated with GnRH 10^{-7} M, galanin 10^{-7} M or neurotensin 10^{-5} M, for 2 h at 37° C. LH in the media was determined by ELISA. Bars represent the means \pm SD's of 8 replicates. * p < 0.001, ** p < 0.005 and *** p < 0.01 vs GnRH alone. $\ddagger p < 0.01$ vs neurotensin alone. Black bars represent basal LH secretion.

RESULTS

Effect of neuropeptides on basal LH release

Basal LH secretion from pituitary cells in the presence or absence of different neuropeptides (including GnRH) was assayed in vitro. Aside GnRH, galanin and neurotensin increased basal LH secretion (Fig 1). The effect of galanin $(10^{-9} \text{ M to } 10^{-5} \text{ M})$ was clearly dose-dependent. The maximal effect elicited by galanin was obtained with the 10⁻⁵ M concentration and was a 3-4 fold increase over the basal secretion (p < 0.001). The effect of neurotensin was also dose-dependent and the maximal effect observed was a two fold increase (p < 0.01). The maximal effects of galanin and neurotensin 10⁻⁵ M represent about 50% and 25-30%, respectively, of the maximal LH release induced by GnRH. Opioid peptides (leucine-enkephalin, methionine-enkephaline, β -endorphin and dynorphin), substance P, neuropeptide Y (NPY) and vasoactive intestinal peptide (VIP) did not significantly alter the basal LH secretion (data not shown).

The neuropeptides that stimulated basal LH secretion were tested on pituitary cells pretreated with the GnRH antagonist Nal-Glu (Fig 2). Nal-Glu 10⁻⁶ M fully suppressed the LH releasing activity of 10^{-7} M GnRH (p < 0.001) or 10^{-5} M neurotensin (p < 0.01), but did not affect the LH releasing activity of 10^{-7} M galanin.

Effect of neuropeptides on GnRH-stimulated LH release

Both leucine-enkephalin and β -endorphin decreased the LH secretion evoked by 10^{-9} M and 10^{-7} M GnRH (Fig 3). Leucine-enkephalin (10^{-9} M to 10^{-5} M) caused a dose-dependent decrease in LH secretion induced by 10^{-9} M and 10^{-7} M GnRH, leading to complete inhibition at 10^{-5} (p < 0.001). Beta-endorphin 10^{-5} M produced about 25% inhibition of LH release (p < 0.05). Figure 4 shows that naloxone prevented the inhibitory action

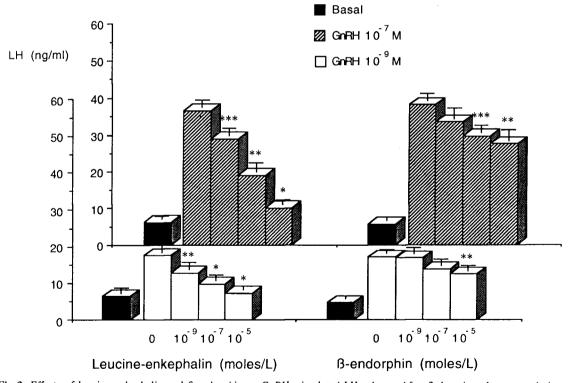


Fig 3. Effects of leucine-enkephalin and β -endorphin on GnRH-stimulated LH release. After 3 days in culture, rat pituitary cells (1.5 x 10⁵ cells/well) were preincubated in the presence of increasing doses of leucine-enkephalin and β -endorphin for 30 min and then were coincubated with GnRH 10^{.9} M or 10^{.7} M for 2 h at 37° C. Media were collected at the end of the experiment and their LH contents determined by ELISA. Bars represent the means \pm SD's of 8 replicates. * p < 0.001, ** p < 0.005 and *** p < 0.05 vs GnRH alone. Black bars represent basal LH secretion.

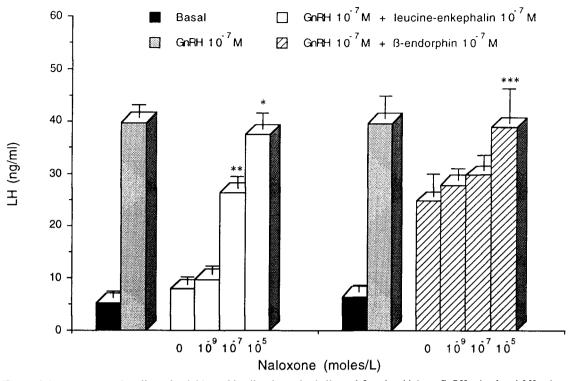


Fig 4. Naloxone reverts the effect of opioid peptides (leucine-enkephalin and β -endorphin) on GnRH-stimulated LH release. Rat pituitary cells (1.5 x 10⁵ cells/well) were cultured for 3 days before the experiment. After 30 min of preincubation in the presence of increasing doses of naloxone, 10 μ l of fresh medium alone or containing leucine-enkephalin or β -endorphin was added and the preincubation continued for 30 min. At the end of the preincubation, cells were coincubated with GnRH 10⁻⁹ M or 10⁻⁷ M for 2 h at 37°C. LH in the medium was determined by ELISA. Bars represent the means \pm SD's of 8 replicates. * p < 0.001, ** p < 0.005 and *** p < 0.01 vs respective control (0). Black bars represent basal LH secretion. Dashed bars represent GnRH-stimulated LH secretion.

of leucine-enkephalin and β -endorphin on GnRH-stimulated LH release (p < 0.001 and 0.01, respectively).

Substance P showed an inverted U doseresponse curve (Fig 5). A dose of 10^{-9} M substance P potentiated the action of GnRH 10^{-9} M (p < 0.001) or 10^{-7} M (p < 0.005). This effect decreased when the dose of substance P was increased to 10^{-7} M and turned inhibitory at 10^{-5} M when 10^{-7} M GnRH was used (p < 0.05). In contrast, neurotensin had no statistically significant effect on GnRHstimulated LH release.

Galanin and NPY enhanced the effect of GnRH (Fig 6). NPY was apparently more effective than galanin in potentiating GnRH effect. Galanin 10^{-7} M and NPY 10^{-9} M doubled the LH released in response to 10^{-9} M GnRH (p < 0.001) and increased 1.5 times the LH release induced by GnRH 10^{-7} M (p < 0.005), respectively.

Vasoactive intestinal peptide, methionineenkephaline and dynorphin had no effect on GnRH-stimulated LH release (data not shown).

DISCUSSION

In previous studies (22), we demonstrated that bovine ME contains an LH-releasing activity not attributable to GnRH. Here, several neuropeptides known to be present in the hypothalamus were tested under the same conditions, to determine which one can account for this activity. Present results show that galanin mimics the effect of ME extracts on LH release since it increased basal as well as GnRH-stimulated LH secretion from rat pituitary cell cultures and was able to stimulate LH release when GnRH receptors were blocked. None of the other neuropeptides tested met all the above criteria, and galanin alone could account for the effects of ME extracts not attributable to GnRH. Furthermore, the maximal response to ME extracts

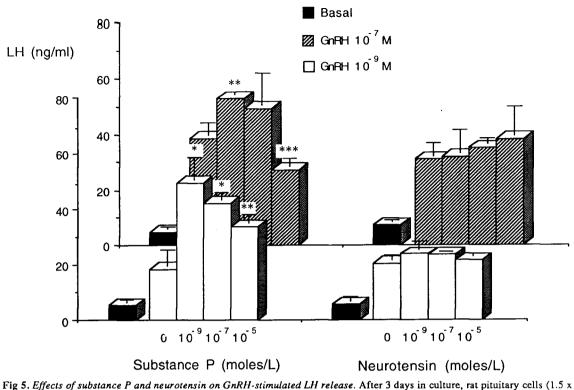


Fig 5. Effects of substance P and neurotensin on GnRH-stimulated LH release. After 3 days in culture, rat pituitary cells (1.5 x 10^5 cells/well) were preincubated in the presence of increasing doses of substance P and neurotensin, for 30 min, and then were coincubated with GnRH 10^9 M or 10^7 M, for 2 h at 37° C. Media were collected at the end of the experiment and their LH contents determined by ELISA. Bars represent the means \pm SD's of 8 replicates. * p < 0.001, ** p < 0.005 and *** p < 0.05 vs GnRH alone. Black bars represent basal LH secretion.

was larger than that to GnRH alone and this difference was suppressed by the GnRH antagonist Nal-Glu (22). Present results show synergism between GnRH and galanin, and unabated effect of the latter in the presence of Nal-Glu. Thus, maximal LH secretory response to ME extracts is best explained by the presence of both GnRH and galanine in the extracts.

Galanin receptors have not been characterized in the anterior pituitary gland. Our results with the GnRH antagonist, Nal-Glu, indicate that galanin effects on LH secretion are neither mediated by the GnRH receptor nor require its cooperation. The effect of neurotensin was peculiar in that it increased basal but not stimulated secretion. The action of neurotensin could involve GnRH receptors because it was blocked by Nal-Glu, but the participation of specific neurotensin receptors sensitive to Nal-Glu is not discarded. Nonetheless, the first interpretation explains better why it acts on basal and not in stimulated secretion. The effects of β-endorphin and leucine-enkephalin were blocked by the opiate antagonist naloxone. This indicates that they modulate LH secretion through specific receptors for opioid peptides that have been found in pituitary gland membranes (19, 23). The effects of NPY and substance P are likely to be mediated by their specific receptors found also in pituitary gland membranes (18, 28, 45).

A striking feature that arises from the comparison of the effect of neuropeptides upon LH secretion in the same assay system is the diversity of responses exhibited by the gonadotrophs. The mechanisms underlying the effect of neuropeptides on LH release have not been fully clarified, but it is conceivable that they may alter diverse links in the chain of events that start with GnRH binding to its receptor and end with the extrusion of the secretory granules. For example, opioid agonist or antagonist produce changes in the number of GnRH receptors (2), phorbol esters up-regulate GnRH receptors (17) and NPY produces an allosteric in-

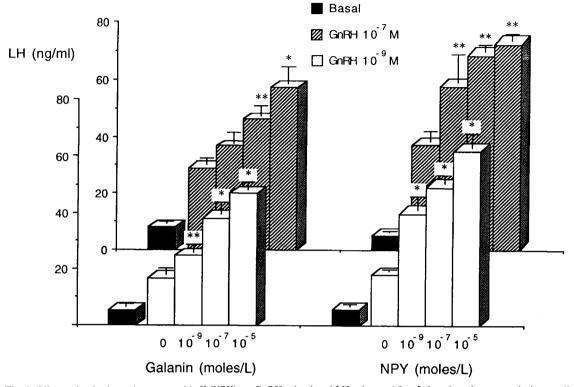


Fig 6. Effects of galanin and neuropeptide Y (NPY) on GnRH-stimulated LH release. After 3 days in culture, rat pituitary cells $(1.5 \times 10^5 \text{ cells/well})$ were preincubated in the presence of increasing doses of galanin and NPY, for 30 min, and then were coincubated with GnRH 10⁻⁹ M or 10⁻⁷ M, for 2 h at 37° C. Media were collected at the end of the experiment and their LH contents determined by ELISA. Bars represent the means \pm SD's of 8 replicates. * p < 0.001, ** p < 0.005 and *** p < 0.05 vs GnRH alone. Black bars represent basal LH secretion.

crease in GnRH binding to its receptor (33). The activation of intracellular messenger systems, such as adenylate cyclase and cAMP-dependent (39) or calcium/phospholipid-dependent protein kinases (40) also augment LH release in response to GnRH or other secretagogues under conditions in which basal hormone release is little affected. The diversity of responses to different neuropeptides under identical conditions, as reported here, offers great potential as a model to investigate the intracellular mechanisms involved in the control of LH release.

The present report adds support to the physiological relevance of neuropeptides in the regulation of LH secretion because the concentrations that were effective are comparable to intrapituitary and/or pituitary portal blood concentrations reported for galanin (24, 32), neurotensin (10), NPY (25, 31), substance P (28) and β -endorphin (12, 44). Previous evidence that supports a physiolog-

ical role of these neuropeptides is that the administration of antisera against ß-endorphin (11), or NPY (13), or substance P (9) or neurotensin (42) modified plasma LH levels. The demonstration of direct effects of several neuropeptides at physiological concentrations supports the concept that gonadotrophs are regulated by a more diversified control system than previously recognized. The most interesting example of pituitary control by multiple factors is the case of corticotrophs, in which the primary regulatory action of corticotrophin-releasing hormone (35) is potentiated and/or supplemented by the stimulatory effects of oxytocin (14) and vasopressin (15) and by catecholamines and angiotensin II (15).

Some of our results are not in full agreement with previous reports (5, 7, 20, 24, 26, 36, 43, 45) and this may be explained by methodological differences. For example, dispersed pituitary cells maintained in culture increase their rate of hormone secretion 5- to 20-fold respect to hemipituitaries (41). The method of cell dispersion, composition of the culture medium and cell density have distinct effects on cell functioning (4), and responsiveness to GnRH changes according to the age and sex of immature rats (34, 37, 38). The importance of these variables makes essential to compare the effects of neuropeptides under identical experimental conditions.

In summary, under the conditions used in this study, opioid peptides are preferentially inhibitory, while galanin, substance P, NPY and neurotensin are preferentially stimulatory on LH secretion. Galanin may account for that portion of the LH-releasing activity of ME extracts not explained by GnRH. This is based on the present observations that galanine increased basal and GnRH-stimulated LH secretion, effects which were not blocked by Nal-Glu. This action of galanin is thus not mediated by the GnRH receptor. The ability of different neuropeptides to modulate LH release at physiological concentrations indicates that regulatory signals complementary to the GnRH system are available for the hypothalamic control of reproductive function. The gonadotrophs in turn offer a wide variety of responses to these neuropeptides and their combinations. Altogether, the data lend further support to a role of galanin in the control of LH secretion.

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